



The evaluation of COVID-19 effect on pregnancy loss; a molecular and diagnostic approach

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Abstract

The severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) virus originates from Wuhan, China; it has spread around the world. According to studies, the results of the SARS-CoV-2 test have been reported positive for some pregnant women. However, not much is known about the effect of this virus on pregnancy and the outcome of baby. The aim of this review study was to evaluate the molecular and diagnostic approach in evaluating the effect of coronavirus disease 2019 (COVID-19) on pregnancy loss. The entry of COVID-19 virus into the pregnant mother's body through various channels, such as the angiotensin-converting enzyme receptor (ACE2) affects immune and coagulation systems and hormone levels. These changes include D-Dimer, platelets and antithrombin III (AT-III) raises and protein C (PC) decrement and also elevated levels of pro-inflammatory cytokines, including IL-6, followed by disruption of various signaling pathways such as JAK / STAT and PI3K. Additionally, decreased expression of cyclooxygenase 1 (COX1) and prostaglandin E2 (PGE2) and hormones such as progesterone were observed. These changes ultimately lead to serious pregnancy risks, including miscarriage. Therefore, identifying pathways by which COVID-19 impairs immune and coagulation systems of pregnant women can be a way to design abortion preventive strategies.

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Introduction

Coronavirus disease 2019 (COVID-19) belongs to the coronavirus family, whose study of clinical effects has become one of the main topics of research in the scientific communities. The pandemic of the virus began in late 2019 in Wuhan, China (1). Severe acute respiratory syndrome coronavirus 2 (SARS-COV-2) is the name now given to this new virus. It is named by the international committee of virus taxonomy (2). COVID-19 can infect all age groups, while it is easily transmitted through the respiratory tract. Clinical signs of the virus include shortness of breath, fever, cough, headache, gastrointestinal and heart problems, blood clotting disorders and other complications (3).

Several studies have focused on the effects of COVID-19 on the general population, but not enough research has been conducted to investigate its effects on people with certain conditions, including pregnancy, while pregnant women are considered as vulnerable group (4,5). In one case, placental infection was

Key point

The COVID-19 can affect coagulation factors in pregnant women which lead to abortion. In COVID-19 infected pregnant women, imbalance between hormones, cytokines and factors cause endometrial decidualization, which may lead to abortion.

observed with COVID-19, which may cause preeclampsia and premature termination of pregnancy or abortion while it may also worsen mother's physiological condition (6). A recent study of 116 pregnant women with COVID-19, found clinical findings of preterm delivery and miscarriage; however, no significant association was found between viral infection and reported cases.

In another case, the traces of COVID-19 in placenta and miscarriage for no reason in the second trimester in a 28-year-old mother can bring us closer to understand the relationship between viral placental infection and miscarriage (7). Due to the clinical effects of this virus on various organs



and to improve the control and management of the pandemic and increase the knowledge about the disease pathogenesis, it is necessary to conduct studies about virus effects on different populations in different conditions such as pregnancy (8). Therefore, in the present review study we evaluated the clinical effects of virus on pregnant women and their specific physiological conditions, as well as the relationship between maternal viral infection and abortion.

COVID-19 and coagulation disorder

Coagulation disorders are one of the clinical symptoms in COVID-19 infected individuals, which cause an imbalance in homeostasis. Among the related factors to the coagulation system, elevated D-dimer has been frequently observed in pregnant women with COVID-19 (9). D-dimer is a protein, which increases in the plasma as a result of blood clotting; high levels of D-dimer can be explained by local increase in fibrin formation. Coagulation disorders and increased risk of disseminated intravascular coagulation are the results of D-dimer and inflammatory cytokines increment. They can lead to clot formation and deposition of fibrin in the placenta bloodstream and prevent proper blood supply to placenta, which may lead to abortion.

Antithrombin III (AT-III) has a binding site for thrombin and heparin, which is an important inhibitor of coagulation cascade. AT-III can inhibit serine-activated proteases such as FXa, FIXa, FXIa, and FXIIa (10). It also potentially prevents activation of nuclear factor- κ B (NF- κ B), which is involved in inflammation. When the COVID-19 enters the body, ACE-2 glycosylation facilitates virus entry into the body. In the virus infected pregnant women, who have recurrent miscarriages, increased AT-III levels have been observed (11).

Protein S (PS) and protein C (PC) also have a vitamin K-dependent anticoagulant role and can bind to activated protein C attached to the phospholipid platelet surface and reduce the effect of FXa on FVa in the coagulation cascade. As a result, PS and PC significantly prevent fibrin clotting and formation (12). PS and protein C also impair the expression of thrombomodulin and endothelial PC receptor on the endothelium. Studies have shown that thrombomodulin has anticoagulant function by binding to thrombin and activating protein C. Decreased protein C levels in COVID-19 infected individuals are associated with imbalance in thrombin regulation (13).

COVID-19 entrance to the body is associated with an increase in interleukin-6 (IL-6) and tumor necrosis factor- α (TNF- α), which are linked to insulin resistance, endothelial dysfunction and blood clotting. Insulin resistance is associated with increased plasminogen activator inhibitor-1 (PAI-1) (14). Increasing PAI-1 has a significant effect on fibrinolytic process by increasing platelet count and decreasing prothrombin activity time and thromboplastin activation. Therefore, maintaining

balance in PAI-1, especially in pregnant women with COVID-19 is essential for a successful pregnancy. Platelet lymphocyte ratio (PLR) and neutrophil lymphocyte ratio (NLR) levels also change in COVID-19 infection. Increased PLR and NLR levels are associated with impression on inflammatory factors and reactive oxygen species, which cause lack of coagulation factors and bleeding and miscarriage (15).

It can now be concluded that COVID-19 can affect coagulation factors; it increases D-dimer, platelets and prothrombin time and also decreases protein C and vitamin K levels, especially in pregnant women. It would be a risk for mother and fetus, and finally it may lead to abortion.

COVID-19 and immune system disorder

The immune system plays an important role in the body homeostasis. Its irregularities can trigger systemic immune responses, following rapid spread of virus, which in turn can lead to COVID-19. The spread of viral agents and inflammation can cause tissue damage, which results in cytokines releasing by damaged cells. Due to the activated signaling cascade, these cytokines affect gene expression. High levels of cytokine in association with a variety of infectious conditions are often called cytokine storms, indicating an immune response characterized by release of interferons, ILs, tumor necrosis factors, chemokines and several other mediators (16).

In COVID-19 infected pregnant women, pro-inflammatory cytokines such as IL-6 increase. This factor causes adverse consequences of abortion or abnormal fetal development. Modulation of the mother's immune system during pregnancy may affect the response to infection, particularly viruses. In general, the immune system during pregnancy adapts to the growth of semi-allogeneic fetus, resulting in an altered immune response to infection during pregnancy. In the placental trophoblast cells of women with the abortion history, this response includes increased production of pro-inflammatory cytokines such as interferon-gamma (IFN- γ)/IL-2/IL-6/IL-7/TNF- α /IL-1 and decreased concentration of anti-inflammatory cytokines (17).

In other words, in the inflammatory and stimulated conditions, IL-1, TNF- α and IL-6 also increase, followed by IL-17 increment; this increase in cytokines activates NF- κ B, MAPK, PI3K and JAK/STAT pathways (18). By activating these signaling pathways, serum and urinary concentration of progesterone-induced blocking factor decreases; as a result, by increasing NK cells and TNF production, NK cells target trophoblast cells and cause spontaneous abortion (19).

The proliferation, differentiation and function of NK cells are regulated by direct effect of progesterone and estrogen on intracellular nuclear receptors, or by intermediate cells in the uterus in early pregnancy. In normal pregnancy, lymphocytes are able to produce progesterone and

progesterone induced blocking factor; they increase fetal immunity, inhibit NK cell activity and reduce the risk of miscarriage by increased production of anti-inflammatory cytokines. However, when immune system becomes abnormal due to virus spread, this ability is lost. Natural killer cells spread increases spontaneous abortions (20). Meanwhile, IL-10 modulates resistance to inflammatory stimuli by reducing the expression regulation of pro-inflammatory cytokines, such as IL-1A, IL-12, IL-6 and TNF α (21). In general, inflammatory cytokines increment, especially IL-6 is associated with activation of STAT 3 and STAT 1 in the JAK/stat pathway and imbalance between Treg/Th17 and cellular immunodeficiency. On the other hand, increasing IL-6 disrupts the expression of genes in this pathway (22).

For example, the sprouty 4 (SPRY4) gene is called IFN- γ , which causes STAT1 phosphorylation via the PI3K/AKT pathway. Increased expression of SPRY4 gene inhibits trophoblast proliferation and increases apoptosis (23). Additionally, the IRF1 gene, which in collaboration with STAT1 reduces miR-103, leads to repeated spontaneous abortions by increasing the M1 macrophage activity (24). Accordingly, STAT3 factor increment in large quantities can suppress GP130 through the suppressor of cytokine signaling 3 (SOCS3), which activates the MAPK/PI3k (25). In these pathways, overexpression of TNF- α and matrix metalloproteinase MMP-3 and MMP-9 factors causes apoptosis of trophoblast cells, by altering B-catenin/wnt pathway activity, inhibiting the integrin-linked kinase (ILK) and also by acting on caspases and activating caspase3 (26).

NF- κ B pathway activation and increase affect hypertension. It causes a significant decrease in uterine endometrial immunity and the immunological mechanism of gestational hypertension syndrome. Eventually, these pathways affect estradiol and intracellular hormones and progesterone-induced blocking factor concentration, increasing the risk of miscarriage (27). On the other hand, it has been shown that inflammation in the dental pulp can also affect abortion incidence. It has been shown that inflammation in the dental pulp can stimulate the immune system and produce inflammatory mediators through dysregulation of some miRs (Table 1). Considering that one of the complications of COVID-19 infection is cytokine storm, and since dental pulp is effective in the occurrence of cytokine storm, possibly COVID-19 infection can cause abortion by dental pulp (28).

Besides, stimulatory effect of IL-6 increases miR-223-3p and mir-184. Elevation of mir-184 by targeting WIG1 increases trophoblast cells apoptosis by regulating FAS expression. mir-223-3p increment also has a regulatory effect on FOX1 and APOL1 genes, which are involved in immune cell homeostasis and regulation of inflammation and apoptosis. Finally, they form a complex monitoring network with pro-inflammatory agents and signaling pathways (34).

Due to elevated levels of these inflammatory cytokines in pregnant women undergoing abortion and given that these inflammatory cytokines increase in pregnant women with COVID-19, it can be concluded that these cytokines by activating the signaling pathways and the changes mentioned, cause adverse consequences in pregnancy, such as abortion or abnormal growth and development of fetus.

COVID-19 and endometrial decidualization

Endometrial decidualization, a vital multicellular process for pregnancy progression is one of the first changes that the uterus adapts to. The interactions between pregnancy-related hormones and cytokines produced by embryonic and uterine cells have been identified as an essential event for decidualization (35). In COVID-19 infected pregnant women, due to lack of coordination and balance between cytokines and hormones, a disorder is observed in the decidualization process (36). Endometriosis is one of the most common diseases in pregnant women, which has been partly explained by resistance to progesterone and decreased intracellular progesterone receptor expression in the extra-uterine endometrium. However, overproduction of progesterone can also impair decidualization by inhibiting leukemia inhibitory factor (LIF)/STAT3 (37).

Heart- and neural crest derivatives-expressed transcript 2 is a major factor in the transcription of progesterin-induced human endometrial stromal cells. It also plays a key role in activation and survival of uterine natural killer by regulating IL-15. Heart- and neural crest derivatives-expressed transcript 2 balances the production of vascular endothelial growth factor and placental growth factor, resulting in balance in endometrial stromal cells (38). COVID-19 infected pregnant women are at the risk of miscarriage, due to decreased level of progesterone production (39).

Studies have shown that ACE2, known as a receptor for COVID-19 entry is essential for decidualization

Table 1. Summary of miRs involved in inflammation by dental pulp

miR	Target	Mechanism	Ref.
miR-21	KBTBD7	Cause activation NF- κ B and lead to inflammation	(29)
miR-410	MMP-14	Decreased expression of miR-410 causes increased MMP-14 production and inflammation	(30)
miR-146a	bFGF	Cause production of inflammatory mediators	(31)
Let-7c-5p	DMP1	Down regulation of Let-7c-5p lead to NF- κ B pathway activation and inflammation	(32)
miR-223-3p	DSPP DMP1	Cause stimulate immune response and inflammation	(33)

Abbreviation: bFGF: Basic fibroblast growth factor; DMP1: Dentin matrix protein-1; MMP-14: Matrix Metalloproteinase; DSPP: Dentin sialophosphoprotein.

of endometrial stromal cells. Angiotensin-converting enzyme 2 is increased during the process of primary human endometrial stromal cell development. Due to increase in ACE2 in various tissues, such as ovaries and uterus, a potential risk to the reproductive system and pregnancy was existed (40). In addition to ACE2, other factors affecting human endometrial stromal cells include prolactin (PRL) and insulin-like growth factor binding protein-1 (IGFBP1). They are produced by decidual cells. Prolactin and insulin-like growth factor binding protein-1 are two known factors for decidual cell maturation and proteoglycan decorin. Decidual cell maturation and proteoglycan decorin inhibit human trophoblast regeneration, migration, invasion and differentiation; while, they are needed to regenerate uterine arteries during normal pregnancy. According to studies, some cytokines increment such as IL-11 is associated with changes in the regulation of expression of IGFBP1 and PRL genes and their incensement. This increase is also achieved through the entry of virus to the pregnant women, followed by activation of other cytokines and changes in the regulation of their expression. As a result, there would be the miscarriage risk during pregnancy (41).

Morphogenetic protein-2 (bmp2) has also been shown to increase the production of pro-inflammatory cytokines. It also reduces the regulation of cyclooxygenase 1 (COX1) expression, followed by a decrease in prostaglandin E2 (PGE2), which is disrupted by the signaling pathway of SMAD1/SMAD5/(Alk3) activin receptor-like kinase 3 in endometrial stromal cells. An increase in the amount of pro-inflammatory cytokines without any controlling effects increases the risk of miscarriage for COVID-19 infected pregnant women (42).

According to the studies, sirtuin 1 (SIRT1) can also be introduced as an effective factor to regulate homeostasis and ESC decidualization (43). Sirtuin 1 through regulating the expression of superoxide dismutase2 (SOD2) and nuclear factor erythroid 2-related factor 2 (NRF2), as well as by deacetylation of Forkhead box O1 (FOXO1) makes adjustment homeostasis of reactive oxygen species (ROS) and NAD⁺; since it also enhances cell protection against oxidative stress. Decreased SIRT1 followed by decreased FOXO1 deacetylation and imbalances in ROS and NAD⁺ observed in patients with COVID-19 cause recurrent implantation failure (44). FOXO1 is also considered as an endometrial stromal cells decidualization marker, which acts as a transcriptional regulator of PRL and Insulin (45).

Studies have also shown that the effect of norepinephrine on decidualization can be investigated. Norepinephrine prevents endometrial decidualization by activating the protein kinase C signaling pathway via B-adrenergic receptor regulation (46). Given the changes mentioned, and the imbalance between hormones, cytokines and factors involved in pregnancy in COVID-19 infected pregnant women, we can conclude that the process of endometrial decidualization is disrupted and subsequently

abortion may happen.

Conclusion

Pregnant women are in a state of suppressed immune system due to physiological changes; they are considered as COVID-19 high-risk group, due to susceptibility to infections and mechanical functions. Immune system suppression disrupts the pregnancy process by affecting the profiles of cytokines and various coagulation systems and hormones. These disorders are associated with the risk of miscarriage. We hope this review be useful for pregnancy and neonatal services, seeking to respond to COVID-19.

Authors' contribution

NA, SKM and MY were the principal investigators of the study. MY and ZJ were included in preparing the concept and design. NH and MR revisited the manuscript and critically evaluated the intellectual contents. All authors participated in preparing the final draft of the manuscript, revised the manuscript and critically evaluated the intellectual contents. All authors have read and approved the content of the manuscript and confirmed the accuracy or integrity of any parts of the work.

Conflicts of interest

The authors declare no conflict of interest

Ethical issues

Ethical issues (including plagiarism, data fabrication, double publication) have been completely observed by the authors.

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References

1. Pericàs JM, Hernandez-Meneses M, Sheahan TP, Quintana E, Ambrosioni J, Sandoval E, et al. COVID-19: from epidemiology to treatment. *Eur Heart J*. 2020;41:2092-112. doi: 10.1093/eurheartj/ehaa462.
2. Li H, Liu SM, Yu XH, Tang SL, Tang CK. Coronavirus disease 2019 (COVID-19): current status and future perspectives. *Int J Antimicrob Agents*. 2020;55(5):105951. doi: 10.1016/j.ijantimicag.2020.105951.
3. Guo YR, Cao QD, Hong ZS, Tan YY, Chen SD, Jin HJ, et al. The origin, transmission and clinical therapies on coronavirus disease 2019 (COVID-19) outbreak-an update on the status. *Mil Med Res*. 2020;7:11. doi: 10.1186/s40779-020-00240-0.
4. Juan J, Gil MM, Rong Z, Zhang Y, Yang H, Poon LC. Effect of coronavirus disease 2019 (COVID-19) on maternal, perinatal and neonatal outcome: systematic review. *Ultrasound Obstet Gynecol*. 2020;56:15-27. doi: 10.1002/uog.22088.
5. Soma-Pillay P, Nelson-Piercy C, Tolppanen H, Mebazaa A. Physiological changes in pregnancy. *Cardiovasc J Afr*. 2016;27:89-94. doi: 10.5830/cvja-2016-021.
6. Lu-Culligan A, Chavan AR, Vijayakumar P, Irshaid L, Courchaine EM, Milano KM, et al. SARS-CoV-2 infection in pregnancy is associated with robust inflammatory response at the maternal-fetal interface. *medRxiv [Preprint]*. January 26, 2021. Available from: <https://www.medrxiv.org/content/10.1101/2021.01.25.21250452v1>.
7. Baud D, Greub G, Favre G, Gengler C, Jaton K, Dubruc E, et al. Second-trimester miscarriage in a pregnant woman with SARS-CoV-2 infection. *JAMA*. 2020;323:2198-200. doi: 10.1001/jama.2020.7233.
8. Afshar Y, Gaw SL, Flaherman VJ, Chambers BD, Krakow

- D, Berghella V, et al. Clinical presentation of coronavirus disease 2019 (COVID-19) in pregnant and recently pregnant people. *Obstet Gynecol.* 2020;136:1117-25. doi: 10.1097/aog.0000000000004178.
9. Zou Y, Guo H, Zhang Y, Zhang Z, Liu Y, Wang J, et al. Analysis of coagulation parameters in patients with COVID-19 in Shanghai, China. *Biosci Trends.* 2020;14:285-9. doi: 10.5582/bst.2020.03086.
 10. Yang S, Chen D, Wang J, Qiao R, Cui L. Coagulation status in women with missed abortion [Preprint]. *Res Sq.* 2021. doi: 10.21203/rs.3.rs-171135/v1.
 11. Ceriello A, De Nigris V, Prattichizzo F. Why is hyperglycaemia worsening COVID-19 and its prognosis? *Diabetes Obes Metab.* 2020;22:1951-2. doi: 10.1111/dom.14098.
 12. Amiral J, Seghatchian J. Revisiting the activated protein C-protein S-thrombomodulin ternary pathway: Impact of new understanding on its laboratory investigation. *Transfus Apher Sci.* 2019;58:538-44. doi: 10.1016/j.transci.2019.06.008.
 13. Tiscia GL, Favuzzi G, De Lorenzo A, Cappucci F, Fischetti L, di Mauro L, et al. Reduction of ADAMTS13 levels predicts mortality in SARS-CoV-2 patients. *TH Open.* 2020;4:e203-e6. doi: 10.1055/s-0040-1716379.
 14. Yanai H. Adiposity is the crucial enhancer of COVID-19. *Cardiol Res.* 2020;11:353-4. doi: 10.14740/cr1118.
 15. Biyik I, Albayrak M, Keskin F. Platelet to lymphocyte ratio and neutrophil to lymphocyte ratio in missed abortion. *Rev Bras Ginecol Obstet.* 2020;42:235-9. doi: 10.1055/s-0040-1709693.
 16. Wong YP, Khong TY, Tan GC. The effects of COVID-19 on placenta and pregnancy: what do we know so far? *Diagnostics (Basel).* 2021;11:94. doi: 10.3390/diagnostics11010094.
 17. Han H, Ma Q, Li C, Liu R, Zhao L, Wang W, et al. Profiling serum cytokines in COVID-19 patients reveals IL-6 and IL-10 are disease severity predictors. *Emerg Microbes Infect.* 2020;9:1123-30. doi: 10.1080/22221751.2020.1770129.
 18. Noack M, Miossec P. Selected cytokine pathways in rheumatoid arthritis. *Semin Immunopathol.* 2017;39:365-83. doi: 10.1007/s00281-017-0619-z.
 19. Szekeres-Bartho J, Šučurović S, Mulac-Jeričević B. The role of extracellular vesicles and PIBF in embryo-maternal immune-interactions. *Front Immunol.* 2018;9:2890. doi: 10.3389/fimmu.2018.02890.
 20. Bogdan A, Berta G, Szekeres-Bartho J. PIBF positive uterine NK cells in the mouse decidua. *J Reprod Immunol.* 2017;119:38-43. doi: 10.1016/j.jri.2016.12.001.
 21. Darif D, Hammi I, Kihel A, El Idrissi Saik I, Guessous F, Akarid K. The pro-inflammatory cytokines in COVID-19 pathogenesis: what goes wrong? *Microb Pathog.* 2021;153:104799. doi: 10.1016/j.micpath.2021.104799.
 22. Venkatesan T, Choi YW, Lee J, Kim YK. Pinus densiflora needle supercritical fluid extract suppresses the expression of pro-inflammatory mediators iNOS, IL-6 and IL-1 β , and activation of inflammatory STAT1 and STAT3 signaling proteins in bacterial lipopolysaccharide-challenged murine macrophages. *Daru.* 2017;25:18. doi: 10.1186/s40199-017-0184-y.
 23. Qin S, Zhang Y, Zhang J, Tian F, Sun L, He X, et al. SPRY4 regulates trophoblast proliferation and apoptosis via regulating IFN- γ -induced STAT1 expression and activation in recurrent miscarriage. *Am J Reprod Immunol.* 2020;83:e13234. doi: 10.1111/aji.13234.
 24. Zhu X, Liu H, Zhang Z, Wei R, Zhou X, Wang Z, et al. MiR-103 protects from recurrent spontaneous abortion via inhibiting STAT1 mediated M1 macrophage polarization. *Int J Biol Sci.* 2020;16:2248-64. doi: 10.7150/ijbs.46144.
 25. Liu X, D'Cruz AA, Hansen J, Croker BA, Lawlor KE, Sims NA, et al. Deleting Suppressor of Cytokine Signaling-3 in chondrocytes reduces bone growth by disrupting mitogen-activated protein kinase signaling. *Osteoarthritis Cartilage.* 2019;27:1557-63. doi: 10.1016/j.joca.2019.05.018.
 26. Chen Q, Ni Y, Han M, Zhou WJ, Zhu XB, Zhang AJ. Integrin-linked kinase improves uterine receptivity formation by activating Wnt/ β -catenin signaling and up-regulating MMP-3/9 expression. *Am J Transl Res.* 2020;12:3011-22.
 27. Arck PC. Stress and pregnancy loss: role of immune mediators, hormones and neurotransmitters. *Am J Reprod Immunol.* 2001;46:117-23. doi: 10.1111/j.8755-8920.2001.460201.x.
 28. Shen Z, Kuang S, Zhang Y, Yang M, Qin W, Shi X, et al. Chitosan hydrogel incorporated with dental pulp stem cell-derived exosomes alleviates periodontitis in mice via a macrophage-dependent mechanism. *Bioact Mater.* 2020;5:1113-26. doi: 10.1016/j.bioactmat.2020.07.002.
 29. Song J, Wu Q, Jiang J, Sun D, Wang F, Xin B, et al. Berberine reduces inflammation of human dental pulp fibroblast via miR-21/KBTBD7 axis. *Arch Oral Biol.* 2020;110:104630. doi: 10.1016/j.archoralbio.2019.104630.
 30. Brodzikowska A, Gondek A, Rak B, Paskal W, Pelka K, Cudnoch-Jędrzejewska A, et al. Metalloproteinase 14 (MMP-14) and hsa-miR-410-3p expression in human inflamed dental pulp and odontoblasts. *Histochem Cell Biol.* 2019;152:345-53. doi: 10.1007/s00418-019-01811-6.
 31. Liu L, Shu S, Cheung GS, Wei X. Effect of miR-146a/bFGF/PEG-PEI nanoparticles on inflammation response and tissue regeneration of human dental pulp cells. *Biomed Res Int.* 2016;2016:3892685. doi: 10.1155/2016/3892685.
 32. Yuan H, Zhang H, Hong L, Zhao H, Wang J, Li H, et al. MicroRNA let-7c-5p suppressed lipopolysaccharide-induced dental pulp inflammation by inhibiting dentin matrix protein-1-mediated nuclear factor kappa B (NF- κ B) pathway in vitro and in vivo. *Med Sci Monit.* 2018;24:6656-65. doi: 10.12659/msm.909093.
 33. Huang X, Liu F, Hou J, Chen K. Inflammation-induced overexpression of microRNA-223-3p regulates odontoblastic differentiation of human dental pulp stem cells by targeting SMAD3. *Int Endod J.* 2019;52:491-503. doi: 10.1111/iej.13032.
 34. Wu J, Niu P, Zhao Y, Cheng Y, Chen W, Lin L, et al. Impact of miR-223-3p and miR-2909 on inflammatory factors IL-6, IL-1 β , and TNF- α , and the TLR4/TLR2/NF- κ B/STAT3 signaling pathway induced by lipopolysaccharide in human adipose stem cells. *PLoS One.* 2019;14:e0212063. doi: 10.1371/journal.pone.0212063.
 35. Oestreich AK, Chadchan SB, Popli P, Medvedeva A, Rowen MN, Stephens CS, et al. The autophagy gene Atg16L1 is necessary for endometrial decidualization. *Endocrinology.* 2020;161:bqz039. doi: 10.1210/endo/bqz039.
 36. Sills ES, Wood SH. An experimental model for peri-conceptual COVID-19 pregnancy loss and proposed interventions to optimize outcomes. *Int J Mol Cell Med.* 2020;9:180-7. doi: 10.22088/ijmcm.bums.9.3.180.
 37. Liang YX, Liu L, Jin ZY, Liang XH, Fu YS, Gu XW, et al. The high concentration of progesterone is harmful for endometrial receptivity and decidualization. *Sci Rep.* 2018;8:712. doi: 10.1038/s41598-017-18643-w.
 38. Murata H, Tanaka S, Tsuzuki-Nakao T, Kido T, Kakita-Kobayashi M, Kida N, et al. The transcription factor HAND2 up-regulates transcription of the IL15 gene in human endometrial stromal cells. *J Biol Chem.* 2020;295:9596-605. doi: 10.1074/jbc.RA120.012753.
 39. Cui L, Xu F, Wang S, Li X, Lin H, Ding Y, et al. Pharmacological activation of rev-erba suppresses LPS-induced macrophage M1 polarization and prevents pregnancy loss. *BMC Immunol.* 2021;22:57. doi: 10.1186/s12865-021-00438-4.

40. Wang N, Qin L, Ma L, Yan H. Effect of severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) on reproductive system. *Stem Cell Res.* 2021;52:102189. doi: 10.1016/j.scr.2021.102189.
41. Eyal O, Jomain JB, Kessler C, Goffin V, Handwerger S. Autocrine prolactin inhibits human uterine decidualization: a novel role for prolactin. *Biol Reprod.* 2007;76:777-83. doi: 10.1095/biolreprod.106.053058.
42. Arikawa T, Omura K, Morita I. Regulation of bone morphogenetic protein-2 expression by endogenous prostaglandin E2 in human mesenchymal stem cells. *J Cell Physiol.* 2004;200:400-6. doi: 10.1002/jcp.20031.
43. Li J, Qi J, Yao G, Zhu Q, Li X, Xu R, et al. Deficiency of sirtuin 1 impedes endometrial decidualization in recurrent implantation failure patients. *Front Cell Dev Biol.* 2021;9:598364. doi: 10.3389/fcell.2021.598364.
44. Yang Y, Li W, Liu Y, Sun Y, Li Y, Yao Q, et al. Alpha-lipoic acid improves high-fat diet-induced hepatic steatosis by modulating the transcription factors SREBP-1, FoxO1 and Nrf2 via the SIRT1/LKB1/AMPK pathway. *J Nutr Biochem.* 2014;25:1207-17. doi: 10.1016/j.jnutbio.2014.06.001.
45. Yoshino O, Osuga Y, Hirota Y, Koga K, Yano T, Tsutsumi O, et al. Akt as a possible intracellular mediator for decidualization in human endometrial stromal cells. *Mol Hum Reprod.* 2003;9:265-9. doi: 10.1093/molehr/gag035.
46. Wang J, Tang Y, Wang S, Cui L, Li D, Du M. Norepinephrine exposure restrains endometrial decidualization during early pregnancy. *J Endocrinol.* 2021;248:277-88. doi: 10.1530/joe-20-0479.